INVITEK
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USER MANUAL

Invisorb® Spin Virus RNA Mini Kit

for purification of viral RNA from up to 200 µl serum, plasma, and other cell-free body fluids,cell culture supernatants, rinse liquid from swabs, stool sample suspensions, cells, fresh or frozen tissue samples

CE IVD REF 1040300300 Invitek Molecular GmbH · D-13125 Berlin

Instruction for the Invisorb® Spin Virus RNA Mini Kit

The **Invisorb® Spin Virus RNA Mini Kit** is the ideal tool using Invisorb® technology for the isolation and purification of high quality viral RNA from RNA viruses contained in up to 200 µl serum, plasma, cerebrospinal fluid, other cell free body fluids and cell culture supernatants, swab material, rinse liquid from swabs stool samples (max. 50 mg), cells (1 x 10^6 mammalian cells) and fresh or frozen tissue samples (max. 20 mg) for in vitro diagnostic purposes using a spin-filter format. Fresh or frozen plasma or serum from blood treated with anticoagulants like EDTA or citrate, *but not with heparin*, as well as small samples whole blood can be used.

The kit is neither suitable for isolation of total RNA from whole blood, blood stains, cultured or isolated cells, tissue samples, bacteria, fungi, plants nor for purification of viral DNA.

CE **IVD** Compliance with EU Directive 98/79/EC on *in vitro* medical devices.

Not for *in-vitro* diagnostic use in countries where the EU Directive 98/79/EC on *in vitro* medical devices is not recognized.

Contents

Kit content of the Invisorb® Spin Virus RNA Mini Kit

	250 viral RNA extractions
Catalogue No.	1040300300
Lysis Buffer RV	170 ml
Carrier RNA	for 3 x 2 ml working solution
Proteinase K	for 3 x 2 ml working solution
Binding Solution (fill with 99.7% Isopropanol)	empty bottle (final volume 120 ml)
Wash Buffer R1	80 ml (final volume160 ml)
Wash Buffer R2	2 x 40 ml (final volume 2 x 200 ml)
Elution Buffer R	30 ml
RNase Free Water	3 x 2 ml
RTA Spin Filter Set	5 x 50
RTA Receiver Tubes	15 x 50
Safe-Lock Tubes 2.0 ml	250
Elution Tubes	5 x 50
Manual	1
Initial steps	Add 2 ml ddH_2O to each tube with **Proteinase K**, mix thoroughly until completely dissolving Fill 120 ml 99.7% **Isopropanol** (molecular biologic grade) into the empty bottle Add 2 ml **RNase Free Water** to each tube Carrier RNA. Mix thoroughly until completely dissolving Add 80 ml 96-100% ethanol to each bottle **Wash Buffer R1** Add 160 ml 96-100% ethanol to each bottle **Wash Buffer R2**

Symbols

 Manufacturer

 Lot number

Attention: *Do not combine components of different kits, unless the lot numbers are identical!*

 Catalogue number

 Expiry date

 Consult operating instructions

 Temperature limitation

 Do not reuse

 Humidity limitation

Storage

All buffers and kit contents of the **Invisorb® Spin Virus RNA Mini Kit,** except **dissolved Proteinase K** and **dissolved Carrier RNA** should be stored at room temperature and are stable for at least 12 months.

Room temperature (RT) is defined as range from 15-30°C.

Before every use, make sure that all components have room temperature. If there are any precipitates within the provided solutions solve these precipitates by warming carefully (up to 30°C).

Proteinase K: Dissolved Proteinase K must be stored at 2 - 8 °C for up to two months. For longer storage –20 °C is recommended, freeze-thaw once only. Therefore, the dissolved Proteinase K is stable as indicated on the kit package.

Carrier RNA: Dissolved Carrier RNA must be stored at -20°C. Therefore, the dissolved mix is stable as indicated on the kit package.

Wash Buffers charged with ethanol should be appropriately sealed and stored at room temperature.

Binding Solution charged with isopropanol should be appropriately sealed and stored at room temperature.

Quality control and product warranty

Invitek Molecular warrants the correct function of the **Invisorb® Spin Virus RNA Mini Kit** for applications as described in this manual. Purchaser must determine the suitability of the Product for its particular use. Should any Product fail to perform the applications as described in the manual, Invitek Molecular will check the lot and if Invitek Molecular investigates a problem in the lot, Invitek Molecular will replace the Product free of charge.

Invitek Molecular reserves the right to change, alter, or modify any Product to enhance its performance and design at any time.

In accordance with Invitek Molecular's EN ISO 13485 certified Quality Management System the performance of all components of the **Invisorb® Spin Virus RNA Mini Kit** have been tested separately against predetermined specifications routinely on lot-to-lot to ensure consistent product quality.

If you have any questions or problems regarding any aspects of **Invisorb® Spin Virus RNA Mini Kit** or other Invitek Molecular products, please do not hesitate to contact us. A copy of Invitek Molecular's terms and conditions can be obtained upon request or are presented at the Invitek Molecular webpage www.invitek-molecular.com.

For technical support or further information please contact:
from Germany: +49-(0)30-9489-2901/ 2910
from abroad: +49-(0)30-9489-2907 or contact your local distributor.

Intended use

The **Invisorb® Spin Virus RNA Mini Kit** is the ideal tool for reliable and fast manual isolation and purification of high quality viral RNA from serum, urine, plasma, cerebrospinal fluid, other cell free body fluids and cell culture supernatants, swab material, stool, cells, and tissue samples. For reproducible high yields an appropriate sample storage and quick operation under the rules for RNA operation is essential. The purified viral RNA is ready to use for *in vitro* diagnostic analysis only.

The isolation protocol and all buffers are optimized to assure a high yield as well as a high purity of purified viral RNA. All manual work is reduced to a minimum.

THE PRODUCT IS INTENDED FOR USE BY PROFESSIONALS ONLY, SUCH AS TECHNICIANS, PHYSICIANS AND BIOLOGISTS TRAINED IN MOLECULAR BIOLOGICAL TECHNIQUES. It is designed to be used with any downstream application employing enzymatic amplification or other enzymatic modifications of RNA followed by signal detection or amplification. Any diagnostic results generated by using the sample preparation procedure in conjunction with any downstream diagnostic assay should be interpreted with regard to other clinical or laboratory findings.

To minimize irregularities in diagnostic results, adequate controls for downstream applications should be used.

The kit complies with EU Directive 98/79/EC on in vitro medical devices. However, it is not for in-vitro diagnostic use in countries where the EU Directive 98/79/EC on in vitro medical devices is not recognized.

Product use limitation

The kit is neither validated for use with bone marrow, cultured cells nor for the isolation of total RNA from serum, plasma, blood, tissue or nor for the isolation of viral DNA.

The included chemicals are only useable once.

Differing of starting material or flow trace may lead to inoperability; therefore, neither a warranty nor guarantee in this case will be given, neither implied nor express. The user is responsible to validate the performance of the Invitek Molecular Product for any particular use. Invitek Molecular does not provide for validation of performance characteristics of the Product with respect to specific applications.

Invitek Molecular products may be used e.g. in clinical diagnostic laboratory systems under following conditions:

- If used in the US, based on the condition that the complete diagnostic system of the laboratory has been validated pursuant to CLIA' 88 regulations.
- For other countries based on the condition that the laboratory has been validated pursuant to equivalents according to the respective legal basis.

All Products sold by Invitek Molecular are subject to extensive quality control procedures (according to EN ISO 13485) and are warranted to perform as described herein. Any problems, incidents or defects shall be reported to Invitek Molecular immediately upon detection thereof.

The chemicals and the plastic parts are for laboratory use only; they must be stored in the laboratory and must not be used for purposes other than intended.

The product with its contents is unfit for consumption.

Safety information

When and while working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles!

Avoid skin contact! Adhere to the legal requirements for working with biological material!

For more information, please consult the appropriate material safety data sheets (MSDS). These are available online in convenient and compact PDF format at www.invitek-molecular.com for each Invitek Molecular Product and its components. If buffer bottles are damaged or leaking, WEAR GLOVES, AND PROTECTIVE GOGGLES when discarding the bottles in order to avoid any injuries.

Invitek Molecular has not tested the liquid waste generated by the **Invisorb® Spin Virus RNA Mini Kit** procedures for residual infectious materials. Contamination of the liquid waste with residual infectious materials is highly unlikely, but cannot be excluded completely. Therefore, liquid waste must be considered infectious and be handled and discarded according to local safety regulations.

European Community risk and safety phrases for the components of the **Invisorb® Spin Virus RNA Mini Kit** to which they apply are listed below as follows:

Proteinase K

Danger

H315-319-334-335 P280-P305-P351-P338

Lysis Buffer RV

Warning

H302-H312-H319-H332-H412-P280-P305+P351+P338-EUH032

Wash Buffer R1

Warning

H302-H332-H412-P280-P305+P351+P338-EUH032

H302: Harmful if swallowed.
H312: Harmful in contact with skin.
H315: Causes skin irritation.
H319: Causes serious eye irritation.
H332: Harmful if inhaled.
H334: May cause allergy or asthma symptoms or breathing difficulties if inhaled.
H335: May cause respiratory irritation.
H412: Harmful to aquatic life with long lasting effects.
P280: Wear protective gloves/protective clothing/eye protection/face protection.
P305+P351+P338: IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.
EUH032: Contact with acids liberates very toxic gas.

Emergency medical information can be obtained 24 hours a day from infotrac:

outside of USA: 1 – 352 – 323 – 3500
in USA : 1 – 800 – 535 – 5053

Product characteristics of the Invisorb® Spin Virus RNA Mini Kit

Starting material	Yield	Time
up to 200 µl cell-free body fluids swab material, rinsed liquid from swab cell culture supernatants 1 x 10^6 mammalian cells max. 20 mg tissue sample max. 50 mg stool sample	depends on the sample (storage and source) ***Note:*** The added Carrier RNA will account for most of the eluted RNA. Quantitative RT-PCR is recommended for determination of the viral RNA yield.	20 minutes

The **Invisorb® Spin Virus RNA Mini Kit** provides a fast and efficient way for reliable isolation of high quality viral RNA from RNA viruses found in a diverse range of starting material. The procedure is suitable for use with plasma or serum; either can contain Citrate or EDTA and other samples. Samples can be fresh, lyophilized or frozen, provided they have not been frozen and thawed more than once. The procedure can be used for isolation of viral RNA from a broad range of RNA viruses.

The amount of purified viral RNA in the **Invisorb® Spin Virus RNA Mini Kit** procedure depends on the sample type, the virus titer, sample source, transport, storage, and age.

The **Invisorb® Spin Virus RNA Mini Kit** simplifies viral RNA isolation by combining efficient lyses of the starting material and the inactivation of exogenous and endogenous RNases.

The sample will be lysed in an optimized Lysis Solution and proteins will be degraded during the lyses with Proteinase K. The liberated RNA is bound onto the membrane of the RTA Spin Filter. Contaminants are removed by repeated wash steps and the purified viral RNA can be eluted in a small volume of Elution Buffer R. The isolated viral RNA is ready to use and should be stored at - 80°C.

Yield and quality of isolated viral RNA is suitable for any molecular-diagnostic detection system. The diagnostic tests should be performed according to manufacturer's specifications. Due to the high purity, the isolated viral RNA is ready to use for a broad panel of downstream applications like:

- RNA dot blots,
- cDNA transcription
- *in vitro* translation,
- RT-PCR*,
- TaqMan® analysis and array technologies.

For technical support or further information please contact: **+49 (0)30-9489-2901/ 2907/ 2910 or your local distributor.**

*) The PCR process is covered by US Patents 4,683,195, and 4,683,202 and foreign equivalents owned by Hoffmann-La Roche AG.

Principle and procedure

The **Invisorb® Spin Virus RNA Mini Kit** procedure comprises the following steps:

- lyses of the virus particle
- adjustment of the binding conditions, followed by the transfer of the sample into the RNA binding RTA Spin Filter
- binding of the viral RNA to the membrane of the RTA Spin Filter
- washing of the membrane and elimination of contaminants and ethanol.Elution of highly pure viral RNA from the membrane

Repeated wash steps make sure that contaminations and enzyme inhibitors are efficiently removed and high purified RNA is eluted in **Elution Buffer R** or Rnase free water.

This manual contains 6 protocols.

Lysis

Samples are lysed under denaturing conditions at elevated temperatures. Due to the strong denaturing lyses conditions in the presence of **Proteinase K** and **Lysis Buffer RV** cells are quickly broken and RNases are inactivated simultaneously. The viral RNA is secured. The addition of Carrier RNA is necessary for the enhancement of viral RNA recovery so a very small number of viral RNA molecules will also be purified. **Carrier RNA** also stabilizes nucleic acids in samples with very small nucleic acid concentrations.

Binding viral RNA

After adding **Binding Solution** to optimize the binding of viral RNA to the RTA Spin Filter membrane, the lysate will be applied onto the RTA Spin Filter and the viral RNA is bound to the surface of the RTA Filter membrane as the lysate is drawn through by centrifugation.

Removing residual contaminants

Contaminants are efficiently washed away using **Wash Buffer R1** and **R2**, while the viral RNA remains bound to the membrane of the RTA Spin Filter.

Elution

High quality viral RNA is eluted from the membrane using **Elution Buffer R** (or RNase Free Water). The eluted RNA is ready to use in different subsequent applications.

Sampling and storage of starting material

Best results are obtained using freshly extracted samples. As long as the samples are not shock frosted with liquid nitrogen or are incubated with RNase inhibitors or denaturing reagents, the RNA is not secured. Therefore, it is essential, that samples are immediately flash frozen subsequent to the harvesting by using liquid nitrogen and are stored at -80°C. RNA contained in such deep frozen samples is stable for months. RNA purification should be processed as soon as possible. Samples can also be stored in **Lysis Buffer RV** for 1 h at room temperature, overnight at 4°C, and for long-term storage at –80°C. Storage under deep frozen conditions is recommended.

Serum, plasma, urine, cerebrospinal fluid or other cell free body fluids, as well as cell culture supernatants, swabs, and stool samples can be stored on ice for 1 - 2 hours, for short time (up to 24 h) samples may be stored at -20°C. For long term storage, we recommend freezing samples at –80°C. Multiple thawing and freezing before isolating the viral RNA should be avoided.

Serum and plasma (and other cell free body fluids)

Following centrifugation, plasma or serum from blood treated with anticoagulants like EDTA or citrate, but not with heparin, can be stored at 2–8°C for up to 6 hours. For long-term storage, freezing at –20°C to –80°C in aliquots is recommended. Repeated freezing and thawing cycles must be avoided because denaturation and precipitation of proteins result in a decrease of the virus titer and thereby reduce the yield of the extracted viral RNA. Occurring cryoprecipitates can be pelleted by briefly centrifuging (6.800 x g for 3 min). The cleared supernatant should be removed, without disturbing the pellet, and processed immediately. This step will not reduce viral titers.

Tissue samples (biopsy material or frozen section)

Best results are obtained with fresh material or material that has been immediately flash frozen and stored at –20°C or –80°C. Repeated freezing and thawing of stored samples should be avoided, since this leads to reduced RNA yield. Use of poor quality starting material influences the RNA yield, too. The amount of purified RNA in the **Invisorb® Spin Virus RNA Mini Kit** procedure using up to 20 mg tissue sample, depends on kind of starting material. The thawing process could be proceed, e.g. directly in **Lysis Buffer RV.**

Invitek Molecular will be released of its responsibilities if other sample materials than described in the Intended Use are processed or if the sample preparation protocols are changed or modified.

Important points before starting a protocol

Immediately upon receipt of the Product, inspect the Product and its components as well as the package for any apparent damages, correct quantities and quality. If there are any unconformities you have to notify Invitek Molecular in writing with immediate effect upon inspection thereof. If buffer bottles are damaged, contact the Invitek Molecular Technical Services or your local distributor. In case of liquid spillage, refer to “Safety Information” (see page 7). Do not use damaged kit components, since their use may lead to poor kit performance.

- Always change pipet tips between liquid transfers. To avoid cross-contaminations, we recommend the use of aerosol-barrier pipet tips.
- All centrifugation steps are carried out at room temperature.
- When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles.
- Discard gloves if they become contaminated.
- Do not combine components of different kits unless the lot numbers are identical.
- Avoid microbial contamination of the kit reagents.
- To minimize the risk of infections from potentially infectious material, we recommend working under laminar air-flow until the samples are lysed.
- This kit should only be used by trained personnel.

Important indications

Preparing RNA

When preparing viral RNA, work quickly during the manual steps of the procedure. The Lysis Buffer RV of the **Invisorb® Spin Virus RNA Mini Kit** simplifies viral RNA isolation by combining efficient lyses of the starting material and the inactivation of exogenous and endogenous RNases. Extreme care should be taken to avoid contaminations with RNases when handling Elution Buffer R.

Storing samples
Frozen Serum or plasma samples must not be thawed more than ones. Repeated freeze – thawing leads to denaturation and precipitation of proteins, resulting in reduced viral titers and therefore reduced yields of viral nucleic acids. In addition cryoprecipitate formed during freeze and thawing will clog the RTA Spin Filter membrane.

Adding carrier RNA
Carrier RNA serves two purposes. Firstly, it enhances the binding of viral acids to the RTA Spin Filter membrane, especially if there are very few target molecules in the sample. Secondly, the addition of large amounts of Carrier RNA reduces the chance of viral RNA degradation in the rare event that RNase molecules are not denaturated by the chaotropic salt and detergents in the Lysis Buffer RV. If Carrier RNA is not added to the Lysis Buffer RV, this may lead to reduced viral RNA recovery.
The use of an internal control is recommended when using the **Invisorb® Spin Virus RNA Mini Kit** in combination with diagnostic amplification systems. Internal Control RNA and reconstituted Carrier RNA should be added to the Lysis Buffer RV and mixed thoroughly by inverting the tube 10 times. To avoid foaming, do not vortex.

Eluting viral RNA
For downstream applications, that require small staring volumes, using viral RNA eluted in 40 µl Elution Buffer R may increase assay sensitivity.
The volume of eluate recovered may be up to 5 µl less than the volume of elution buffer applied to the RTA Spin Filter. The volume of eluate recovered depends on the nature of the sample.

Handling of RTA Spin Filter
Due to the sensitivity of viral RNA amplification technologies the following precautions are necessary when handling RTA Spin Filter to avoid cross-contamination between sample preparations:

- carefully apply the sample or solution to the RTA Spin Filter, pipet the sample into the
- filter without wetting the rim of the column
- always change pipet tips between liquid transfers, we recommend the use of aerosol-
- barrier pipet tips
- avoid touching the RTA Spin Filter membrane with the pipet tip

Yield and quality of viral RNA

Different amplification systems vary in efficiency depending on the total amount of nucleic acid present in the reaction. Eluates derived by this kit will contain Carrier-RNA, which will greatly exceed the amount of the isolated NA.
Yields of viral nucleic acids isolated from biological samples are usually low concentrated and therefore almost impossible to determine photometrically*.

* Keep in mind that the Carrier-RNA (5 µg per 200 µl sample) will account for most of the present RNA.

The kit is suitable for downstream analysis with NAT techniques, for examples qPCR, RT qPCR, LAMP, LCR. Diagnostic assays should be performed according to the manufacturer's instructions.

Quantitative RT-PCR is recommended for determination of viral RNA yield.

**) In Gel Electrophoresis and in Capillary Electrophoresis, RNA extracted with the provided kit looks like degraded cause the kit contains Carrier RNA, this is poly A RNA in fragments of 100 up to 1000 bases. The kit is not dedicated for applications using this kind of analysis.*

Internal control (IC) / Extraction control

Internal Controls (IC) from the PCR assay provider can be used as extraction controls if the fragments are longer than 100 bp. In this case, they have to be added after finalization of the lysis step. Alternatively, it can be mixed with the Carrier RNA.

Attention: Do not add directly these Internal Controls to the sample!

Preparing buffers

250 viral RNA-extractions:
Add 2 ml ddH_2O to the tube with **Proteinase K,** mix thoroughly until completely
Fill 120 ml 99.7% **Isopropanol** (molecular biologic grade) into the empty bottle
Add 2 ml **RNase Free Water** to the **Carrier RNA.** Mix thoroughly until completely dissolving.
Add 80 ml 96-100% ethanol to each bottle **Wash Buffer R1.**
Add 160 ml 96-100% ethanol to each bottle **Wash Buffer R2**.

Equipment and reagents to be supplied by user

When working with chemicals, always wear a suitable lab coat, disposable gloves and protective goggles. For more information, please consult the appropriate material safety data sheets (MSDS). (See our webpage: www.invitek-molecular.com)

- Microcentrifuge (≥ 11.000 rpm)
- Thermomixer (65°C-80°C)
- ddH_2O
- Ethanol (96-100%)
- Isopropanol *

*The **Invisorb® Spin Virus RNA Mini Kit** is validated with 2-Propanol; Rotipuran >99.7%, p.a., ACS, ISO (Order no. 6752) from **Carl Roth.**

*** Possible suppliers for Isopropanol:**

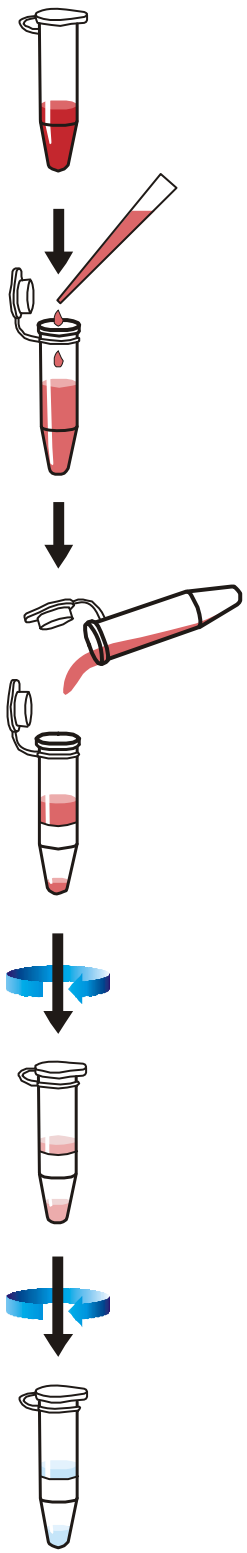
Carl Roth
2-Propanol
Rotipuran >99.7%, p.a., ACS, ISO
Order no. 6752

Applichem
2-Propanol für die Molekularbiologie
Order no. A3928

Sigma
2-Propanol
Order no. 59304-1L-F

Scheme of the Invisorb® Spin Virus RNA Mini Kit

Please work quickly and perform all extraction steps at room temperature (RT)!



Please read the protocols carefully prior to the start of the preparation procedure!

transfer 200 µl sample into a 2.0 ml Safe-Lock Tube
add 600 µl **Lysis Buffer RV,** 20 µl **Carrier RNA**, and 20 µl **Proteinase K** or
or: add 640 µl Mastermix (see page 14)
incubate for 10 minutes at 65°C in a thermomixer

Add the Internal Control to each sample or check page 14**

for realization of the optimal binding conditions add 400 µl **Binding Solution** and mix the sample completely by pipetting up and down or by vortexing

transfer 650 µl of the sample on the RTA Spin Filter, incubate for 1 min and centrifuge for 1 min at 5.900 x g (8.000 rpm)

discard the flow through, transfer the residual sample into the RTA Spin Filter, incubate for 1 min and centrifuge for 1 min at 5.900 x g (8.000 rpm)
transfer the RTA Spin Filter into a new RTA Receiver Tube

pipet 600 µl **Wash Buffer R1** onto the RTA Spin Filter and centrifuge for 1 min at 5.900 x g (8.000 rpm)
discard the flow through and the RTA Receiver Tube
transfer the RTA Spin Filter into a new RTA Receiver Tube

pipet 600 µl **Wash Buffer R2** onto the RTA Spin Filter and centrifuge for 1 min at 5.900 x g (8.000 rpm)
discard the flow through
transfer the RTA Spin Filter into a new RTA Receiver Tube
(Repeat this washing step!)

to eliminate any traces of ethanol, centrifuge again for 1 min at maximum speed
discard the RTA Receiver Tube

transfer the RTA Spin Filter into a RNase free 1.5 ml Elution Tube
pipet 50 - 100 µl of **Elution Buffer R** (preheated to 65°C) directly onto the membrane of the RTA Spin Filter, incubate for 3 min and centrifuge for 1 min at 5.900 x g (8.000 rpm)

discard the RTA Spin Filter and place the eluted viral RNA immediately on ice!

Lysis procedures

For easier handling we recommend to prepare a **Master Mix** only for the needed amount of samples consisting of Lysis Buffer RV, Proteinase K and if required Carrier RNA. When preparing the **Master Mix** it is recommended to use a volume of 5 % greater than that required. The **Master Mix** is stable for at least 2h at RT.

Preparation of a Master Mix

Number of samples	Amount of **Lysis Buffer RV**	Amount of **Carrier RNA**	Amount of **Proteinase K**
	600 µl / sample	20 µl / sample	20 µl / sample
6	3.9 ml	130 µl	130 µl
8	5.1 ml	170 µl	170 µl
10	6.3 ml	210 µl	210 µl
12	7.8 ml	260 µl	260 µl
16	10.2 ml	340 µl	340 µl
20	12.6 ml	420 µl	420 µl
24	15.0 ml	500 µl	500 µl
32	20.1 ml	670 µl	670 µl
40	25.2 ml	840 µl	840 µl
48	30.0 ml	1000 µl	1000 µl

Extraction control

Extraction control DNA or RNA must be combined with the provided Carrier RNA in one mixture. The vials with Carrier RNA contain 240 µl, 1.2 ml or 2.0 ml stock solutions depending on the package size.

Add the respective amount of Extraction Control Nucleic Acid to the Carrier RNA, replace the according amount of RNAse free water. Then you may add the respective mixture to the master mix, **please note that this is just possible, if all the samples need the same control!**

Notes:

If you only have indication of amount per reaction, please calculate by using eluate and template volume.

If the extraction control is stable in plasma, serum, CSF, urine, respiratory samples, whole blood, stool, transport media, or on dried swabs (e.g., armored RNA), it can alternatively be added to the sample shortly before beginning sample preparation.

If the extraction control is naked DNA or RNA, it is unstable in plasma, serum, CSF, urine, respiratory samples, whole blood, stool, transport media, or on dried swabs and must not be added directly to the samples. You may add it together with the carrier RNA, or after finishing the Lysis Step, before continuing with step 2.

Refer to the manufacturer's instructions to determine the optimal amount of extraction control for specific downstream applications. Using an amount other than that recommended may lead to wrong quantification results.

Instructions

The following notes are valid for all protocols:

Note: *The RNA can also be eluted with a lower (but not lower than 40 µl) or a higher volume of Elution Buffer R (depends on the expected yield or needed concentration of RNA).*

Important *After extraction, place the Elution Tube on ice. For a long time storage, place the nucleic acids at –20°C or –80°C.*

Note: *The centrifugation steps were made with the **Centrifuge 5415 D from Eppendorf.** The indicated **rpm amounts** are referring to this centrifuge.*

Protocol 1: Isolation of viral RNA from serum, plasma or other cell-free body fluids

Please read the instructions carefully and conduct the prepared procedure.

Important *Preheat the prospective volume of the **Elution Buffer R** to 80°C for the final elution step.*

The protocol has been optimized for the isolation of RNA from body fluids of 200 µl. For samples, which have a smaller volume than 200 µl, fill up to a total volume of 200 µl with ddH_2O.

1. Sample Lysis

Mix samples smaller than 200 µl with ddH_2O to a total volume of 200 µl. Transfer the sample in a 2.0 ml Safe-Lock Tube. Add 600 µl **Lysis Buffer RV**, 20 µl **Carrier RNA** and 20 µl **Proteinase K** to the sample. Close the cap and vortex shortly. Place the Safe-Lock into a thermomixer and incubate under continuously shaking at 65°C for 10 min.

Add the Internal Control

2. Binding of the RNA

Add 400 µl **Binding Solution** to the tube with the lysed sample and mix the sample completely by pipetting up and down or by vortexing. Transfer 650 µl of the sample into the RTA Spin Filter Set. Close the tube, incubate for 1 min at RT and centrifuge for 1 minute at 5.900 x g (8.000 rpm). Discard the filtrate and transfer the residual sample into the RTA Spin Filter Set. Close the cap, incubate for 1 min at RT and centrifuge at 5.900 x g (8.000 rpm) for 1 min. Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

3. First Washing of the RTA Spin Filter

Add 600 µl **Wash Buffer R1** to the RTA Spin Filter and centrifuge for 1 min at 5.900 x g (8.000 rpm). Discard the filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

4. Second Washing of the RTA Spin Filter

Add 600 µl **Wash Buffer R2** to the RTA Spin Filter and centrifuge for 1 min at 5.900 x g (8.000 rpm). Discard the filtrate, place the RTA Spin Filter again into a new RTA Receiver Tube and repeat the second washing step. Remove the residual ethanol by final centrifugation for 4 min at maximum speed.

5. Elution of the RNA

Place the RTA Spin Filter into a 1.5 ml Elution Tube and add 100 µl of the **Elution Buffer R** (preheated to 80°C) directly onto the RTA Spin Filter surface. Incubate for 3 min. and centrifuge at 5.900 x g (8.000 rpm) for 1 min.

Protocol 2: Isolation of viral RNA from swab material

Please read the instructions carefully and conduct the prepared procedure.

Important *Preheat the prospective volume of the **Elution Buffer R** to 80°C for the final elution step.*

If the swab is delivered in a stabilization media, make sure that these media are suitable for RNA virus stabilization. If you are not sure, please contact the manufacturer.

Some RNA virus stabilizing media dissolve the viral particles. They may be used by transferring 200 µl directly into the 2.0 ml Safe-Lock Tubes and continue with step 1.

1. Sample Lysis

A) If you get a swab without transport media, please follow the instructions below:
- rinse each swab with 500 µl cooled water or cooled PBS and use a 200 µl aliquot of the liquid for viral RNA extraction.
- transfer each sample in a 2.0 ml Safe-Lock Tube and follow the protocol

B) If you want use the swab direct: place the swab in a 2.0 ml Safe-Lock Tube and add 200 µl ddH_2O.

Add 600 µl **Lysis Buffer RV**, 20 µl **Carrier RNA** and 20 µl **Proteinase K** to the sample. Close the cap and vortex shortly. Place the Safe-Lock Tube into a thermomixer and incubate under continuously shaking for 10 min. at 65°C.

Add the Internal Control

Important Note*: To get maximum yield of viral nucleic acids it is essential to leave the swab during the complete lysis time into the reaction tube. It is possible to cut the shaft of the swab, so that you can close the cap of the tube. The removing of the swab from the tube ahead of time will be lead to a dramatically reduced final yield! After lysis time, carefully squeeze out the swab on the wall of the tube and discard the swab.*

2. Binding of the RNA

Add 400 µl **Binding Solution** to the tube with the lysed sample and mix the sample completely by pipetting up and down or by vortexing. Transfer 650 µl of the sample into the RTA Spin Filter Set. Close the tube, incubate for 1 min at RT and centrifuge for 1 minute at 5.900 x g (8.000 rpm). Discard the filtrate and transfer the residual sample into the RTA Spin Filter Set. Close the cap, incubate for 1 min at RT and centrifuge at 5.900 x g (8.000 rpm) for 1 min. Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube

3. First Washing of the RTA Spin Filter

Add 600 µl **Wash Buffer R1** to the RTA Spin Filter and centrifuge for 1 min at 5.900 x g (8.000 rpm). Discard the filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

4. Second Washing of the RTA Spin Filter

Add 600 µl **Wash Buffer R2** to the RTA Spin Filter and centrifuge for 1 min at 5.900 x g (8.000 rpm). Discard the filtrate, place the RTA Spin Filter again into a new RTA Receiver Tube and repeat the second washing step. Remove the residual ethanol by final centrifugation for 4 min at maximum speed.

5. Elution of the RNA

Place the RTA Spin Filter into a 1.5 ml Elution Tube and add 100 µl of the **Elution Buffer R** (preheated to 80°C) directly onto the RTA Spin Filter surface. Incubate for 3 min. and centrifuge at 8.000 rpm for 1 min.

Protocol 3: Isolation of viral RNA from cell culture supernatants

Please read the instructions carefully and conduct the prepared procedure.

Important *Preheat the prospective volume of the **Elution Buffer R** to 80°C for the final elution step.*

1. Sample Lysis

Transfer 200 µl of the cell culture supernatant (cell culture media) into a 2.0 ml Safe-Lock Tube. Add 600 µl **Lysis Buffer RV**, 20 µl **Carrier RNA** and 20 µl **Proteinase K** to the sample. Close the cap and vortex shortly. Place the tube into a thermomixer and incubate under continuously shaking for 10 minutes at 65°C.

Add the Internal Control.

2. Binding of the RNA

Add 400 µl **Binding Solution** to the tube with the lysed sample and mix the sample completely by pipetting up and down or by vortexing. Transfer 650 µl of the sample into the RTA Spin Filter Set. Close the tube, incubate for 1 min at RT and centrifuge for 1 minute at 5.900 x g (8.000 rpm). Discard the filtrate and transfer the residual sample into the RTA Spin Filter Set. Close the cap, incubate for 1 min at RT and centrifuge at 5.900 x g (8.000 rpm) for 1 min. Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

3. First Washing of the RTA Spin Filter

Add 600 µl **Wash Buffer R1** to the RTA Spin Filter and centrifuge for 1 min at 5.900 x g (8.000 rpm). Discard the filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

4. Second Washing of the RTA Spin Filter

Add 600 µl **Wash Buffer R2** to the RTA Spin Filter and centrifuge for 1 min at 8.000 rpm. Discard the filtrate, place the RTA Spin Filter again into a new RTA Receiver Tube and repeat the second washing step. Remove the residual ethanol by final centrifugation for 4 min at maximum speed.

5. Elution of the RNA

Place the RTA Spin Filter into a 1.5 ml Elution Tube and add 100 µl of the **Elution Buffer R** (preheated to 80°C) directly onto the RTA Spin Filter surface. Incubate for 3 min. and centrifuge at 5.900 x g (8.000 rpm) for 1 min.

Protocol 4: Isolation of viral RNA from 1 x 10^6 mammalian cells

Please read the instructions carefully and conduct the prepared procedure.

Important *Preheat the prospective volume of the **Elution Buffer R** to 80°C for the final elution step.*

Harvest cells

Cells grown in suspension: Spin up to 1 x 10^6 cells for 5 min at 1.500 rpm. Discard the supernatant and remove all media completely.

Cells grown in a monolayer: In large culture vessels (dishes > ∅ 35 mm, flasks > 12.5 cm^2) detach cells by trypsination. Transfer the cells to a centrifuge tube and sediment by centrifugation at 1.500 rpm for 5 min. Remove the supernatant completely. In small culture vessels (96-, 24-, 12-, 6-well plates, ∅ 35 mm dishes, 12.5 cm^2 flasks) discard the media completely and continue with the lysis immediately.

Important: *Incomplete removal of the cell culture media will inhibit the lysis and dilute the lysate, which will affect the binding of RNA to the RTA Spin Filter.*

Disrupt cells by adding Lysis Buffer RV

For pelleted cells: Loosen cell pellet by flicking the tube and add 600 µl **Lysis Buffer RV**, 200 µl ddH_2O, 20 µl **Carrier RNA** and 20 µl **Proteinase K**. Close the cap and vortex shortly. No cell clumps should be visible before proceeding with the next step. Pipet the lysed mixture into a 2.0 ml reaction tube.

For monolayer cells: Add 600 µl **Lysis Buffer RV**, 200 µl ddH_2O, 20 µl **Carrier RNA** and 20 µl **Proteinase K** to monolayer cells. Collect cell lysate with a rubber policeman. Mix thoroughly by pipetting up and down. No cell clumps should be visible before proceeding with the next step. Pipet the lysed mixture into 2.0 ml reaction tube.

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1. Sample Lysis

Place the tube with the sample into a thermomixer and incubate under continuously shaking for 10 minutes at 65°C.

Add the Internal Control

2. Binding of the RNA

Add 400 µl **Binding Solution** to the tube with the lysed sample and mix the sample completely by pipetting up and down or by vortexing. Transfer 650 µl of the sample into the RTA Spin Filter Set. Close the tube, incubate for 1 min at RT and centrifuge for 1 minute at 5.900 x g (8.000 rpm). Discard the filtrate and transfer the residual sample into the RTA Spin Filter Set. Close the cap, incubate for 1 min at RT and centrifuge at 5.900 x g (8.000 rpm) for 1 min. Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

3. First Washing of the RTA Spin Filter

Add 600 µl **Wash Buffer R1** to the RTA Spin Filter and centrifuge for 1 min at 5.900 x g (8.000 rpm). Discard the filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

4. Second Washing of the RTA Spin Filter

Add 600 µl **Wash Buffer R2** to the RTA Spin Filter and centrifuge for 1 min at 5.900 x g (8.000 rpm). Discard the filtrate, place the RTA Spin Filter again into a new RTA Receiver Tube and repeat the second washing step. Remove the residual ethanol by final centrifugation for 4 min at maximum speed.

5. Elution of the RNA

Place the RTA Spin Filter into a 1.5 ml Elution Tube and add 100 µl of the Elution Buffer R (preheated to 80°C) directly onto the RTA Spin Filter surface. Incubate for 3 min. and centrifuge at 5.900 x g (8.000 rpm) for 1 min.

Protocol 5: Isolation of viral RNA from max. 20 mg tissue samples

Please read the instructions carefully and conduct the prepared procedure.

Important *Preheat the prospective volume of the **Elution Buffer R** to 80°C for the final elution step.*

Disruption and lysis of the starting material

Note: *To maximize the final yield of viral RNA a complete disruption of tissue sample is important! For the disruption of starting material, it is possible to use commercially available rotor-stator homogenizer or bead mills. It is also possible to disrupt the starting material using mortar and pestle in liquid nitrogen and grind the tissue sample to a fine powder.*

○ ***using rotor-stator homogenizer***

1. Transfer the weighed amount of fresh or frozen starting material in a suitable reaction vesicle for the homogenizer.
2. Add 600 µl **Lysis Buffer RV** and 200µl ddH_20 (vigorously mixed before adding).
3. Homogenize the sample.
4. Transfer the sample into a 2.0 ml reaction tube and place the homogenate for longer storage at –20°C or use the sample immediately for isolation of viral RNA following the protocol step 1.

○ ***using a mortar and pestle and liquid nitrogen***

1. Transfer the weighed amount of fresh or frozen starting material under liquid nitrogen and grind the material to a fine tissue powder.
2. Transfer the powder into a 2.0 ml Safe-Lock Tube. Do not allow the sample to thaw!
3. Add 600 µl **Lysis Buffer RV** and 200 µl ddH_20 to the sample and follow protocol step 1.

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1. Sample Lysis
Add 20 µl **Carrier RNA** and 20 µl **Proteinase K** to the homogenate. Close the cap and vortex shortly. Place the Safe-Lock Tube into a thermomixer and incubate under continuously shaking for 10 minutes at 65°C.

Add the Internal Control

2. Binding of the RNA
Add 400 µl **Binding Solution** to the tube with the lysed sample and mix the sample completely by pipetting up and down or by vortexing. Transfer 650 µl of the sample into the RTA Spin Filter Set. Close the tube, incubate for 1 min at RT and centrifuge for 1 minute at 5.900 x g (8.000 rpm). Discard the filtrate and transfer the residual sample into the RTA Spin Filter Set. Close the cap, incubate for 1 min at RT and centrifuge at 5.900 x g (8.000 rpm) for 1 min. Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

3. First Washing of the RTA Spin Filter
Add 600 µl **Wash Buffer R1** to the RTA Spin Filter and centrifuge for 1 min at 5.900 x g (8.000 rpm). Discard the filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

4. Second Washing of the RTA Spin Filter
Add 600 µl **Wash Buffer R2** to the RTA Spin Filter and centrifuge for 1 min at 5.900 x g (8.000 rpm). Discard the filtrate, place the RTA Spin Filter again into a new RTA Receiver Tube and repeat the second washing step. Remove the residual ethanol by final centrifugation for 4 min at maximum speed.

5. Elution of the RNA
Place the RTA Spin Filter into a 1.5 ml Elution Tube and add 100 µl of the **Elution Buffer R** (preheated to 80°C) directly onto the RTA Spin Filter surface. Incubate for 3 min. and centrifuge at 5.900 x g (8.000 rpm) for 1 min.

Protocol 6: Isolation of viral RNA from max. 50 mg stool samples

Please read the instructions carefully and conduct the prepared procedure.

Important *Preheat the prospective volume of the **Elution Buffer R** to 80°C for the final elution step.*

Sample Preparation

1. Add for each stool sample 400 µl ddH_2O into a 2.0 ml Safe-Lock Tube.
2. Transfer with a new glass stick from each stool sample a small piece (size of a lentil) in the water prefilled 2.0 ml Safe-Lock Tube.
3. Resuspend the sample from the glass stick in water and discard the stick.
4. Close the tube and vortex each sample vigorously until it is a homogeneous suspension
5. Centrifuge the samples for 5 min at max. speed (e.g. at 15.000 rpm Hettich Universal 30 RF)
6. Pipette from each sample 200 µl supernatant and transfer the sample in a new 2.0 ml Safe-Lock Tubes.
7. Add 600 µl **Lysis Buffer RV** to this supernatant and mix vigorously.

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1. Sample Lysis
Add 20 µl **Carrier RNA** and 20 µl **Proteinase K** to the homogenate. Close the cap and vortex shortly. Place the Safe-Lock Tube into a thermomixer and incubate under continuously shaking for 10 minutes at 65°C.

Add the Internal Control.

2. Binding of the RNA
Add 400 µl **Binding Solution** to the tube with the lysed sample and mix the sample completely by pipetting up and down or by vortexing. Transfer 650 µl of the sample into the RTA Spin Filter Set. Close the tube, incubate for 1 min at RT and centrifuge for 1 minute at 5.900 x g (8.000 rpm). Discard the filtrate and transfer the residual sample into the RTA Spin Filter Set. Close the cap, incubate for 1 min at RT and centrifuge at 5.900 x g (8.000 rpm) for 1 min. Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

3. First Washing of the RTA Spin Filter
Add 600 µl **Wash Buffer R1** to the RTA Spin Filter and centrifuge for 1 min at 5.900 x g (8.000 rpm). Discard the filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

4. Second Washing of the RTA Spin Filter
Add 600 µl **Wash Buffer R2** to the RTA Spin Filter and centrifuge for 1 min at 5.900 x g (8.000 rpm). Discard the filtrate, place the RTA Spin Filter again into a new RTA Receiver Tube and repeat the second washing step. Remove the residual ethanol by final centrifugation for 4 min at maximum speed.

5. Elution of the RNA
Place the RTA Spin Filter into a 1.5 ml Elution Tube and add 100 µl of the **Elution Buffer R** (preheated to 80°C) directly onto the RTA Spin Filter surface. Incubate for 3 min. and centrifuge at 5.900 x g (8.000 rpm) for 1 min.

Protocol 7: Isolation of viral RNA from whole blood

Please read the instructions carefully and conduct the prepared procedure.

Important *Preheat the prospective volume of the **Elution Buffer R** to 80°C for the final elution step.*

The protocol has been optimized for the isolation of RNA from 50 µl whole human blood (EDTA or citrate). Please fill up each blood sample to a total volume of 200 µl with ddH_2O or PBS buffer.

Important ***Animal Blood****: The useable volume of the blood depend on animal species. This is to check for every species*

1. Sample Lysis

Transfer the sample in a 2.0 ml Safe-Lock Tube. Add 600 µl **Lysis Buffer RV**, 20 µl **Carrier RNA** and 20 µl **Proteinase K** to the sample. Close the cap and vortex shortly. Place the Safe-Lock Tube into a thermomixer and incubate under continuously shaking at 65°C for 10 min.

Add the Internal Control.

2. Binding of the RNA

Add 400 µl **Binding Solution** to the tube with the lysed sample and mix the sample completely by pipetting up and down or by vortexing. Transfer 650 µl of the sample into the RTA Spin Filter Set. Close the tube, incubate for 1 min at RT and centrifuge for 1 minute at 5.900 x g (8.000 rpm). Discard the filtrate and transfer the residual sample into the RTA Spin Filter Set. Close the cap, incubate for 1 min at RT and centrifuge at 5.900 x g (8.000 rpm) for 1 min. Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

3. First Washing of the RTA Spin Filter

Add 600 µl **Wash Buffer R1** to the RTA Spin Filter and centrifuge for 1 min at 5.900 x g (8.000 rpm). Discard the filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

4. Second Washing of the RTA Spin Filter

Add 600 µl **Wash Buffer R2** to the RTA Spin Filter and centrifuge for 1 min at 5.900 x g (8.000 rpm). Discard the filtrate, place the RTA Spin Filter again into a new RTA Receiver Tube and repeat the second washing step. Remove the residual ethanol by final centrifugation for 4 min at maximum speed.

5. Elution of the RNA

Place the RTA Spin Filter into a 1.5 ml Elution Tube and add 100 µl of the **Elution Buffer R** (preheated to 80°C) directly onto the RTA Spin Filter surface. Incubate for 3 min. and centrifuge at 5.900 x g (8.000 rpm) for 1 min.

Troubleshooting

Problem/ probable cause	Comments and suggestions
Clogged RTA Spin Filter	
Insufficient disruption or homogenization of the starting material.	After lysis spin lysate to pellet debris and continue with the protocol using the supernatant. Increase g-force and/ or centrifugation time. Reduce the amount of starting material. All centrifugation steps should be conducted at room temperature.
Little or no viral RNA eluted	
Incomplete removal of cell culture medium.	Make sure that the cell culture medium is complete removed after the cell harvest.
Insufficient mixing of the sample with Binding Solution	Mix sample sufficient by pipetting up and down with Binding Solution prior to transfer the sample onto the RTA Spin Filter.
Incomplete elution.	Prolong the incubation time with preheated **Elution Buffer R** to 5-10 min or repeat elution step once again.
RNA degraded	
Inappropriate handling of the starting material.	The RNA purification protocol should be performed quickly (see also “General notes on handling RNA”, page 22). Cell pellets stored at - 80°C for later processing should be immediately frozen after cell harvest by liquid nitrogen treatment.
Viral RNA does not perform well in downstream-applications (e.g. RT-PCR)	
Ethanol carryover during elution.	Increase g-force or centrifugation time when drying the RTA Spin Filter.
Salt carryover during elution.	Ensure that **Wash Buffer R1** and **R2** are at room temperature. Check up **Wash Buffer R1** and **R2** for salt precipitates. If there are any precipitates solve these precipitates by careful warming.

Appendix

General notes on handling RNA

RNA is far less stable than DNA. It is very sensitive to degradation by endogenous RNases in the biological material and exogenous RNases which are permanently present everywhere in the lab. To achieve satisfactory qualitative and quantitative results in RNA preparations, contaminations with exogenous RNases has to be reduced as much as possible. Avoid handling bacterial cultures, cell cultures or other biological sources of RNases in the same lab where the RNA purification is to be carried out.

All glassware should be treated before use to ensure that it is RNase-free. Glassware should be cleaned with detergent, thoroughly rinsed and oven baked at 240°C for four or more hours before use. Autoclaving alone will not completely inactivate many RNases. Oven baking will both inactivate RNases and ensure that no other nucleic acids (such as Plasmid DNA) are present on the surface of the glassware. You can also clean glassware with 0.1% DEPC (diethyl pyrocarbonate). The glassware have to stand 12 hours at 37°C and then autoclave or heat to 100°C for 15 min to remove residual DEPC.

- Electrophoresis tanks should be cleaned with detergent solution (e.g. 0.5% SDS), thoroughly rinsed with Rnase free water, and then rinsed with ethanol and allowed to dry.
- Non-disposable plastic ware should be treated before use to ensure that it is Rnase free. Plastic ware should be thoroughly rinsed with 0.1 M NaOH, 1 mM EDTA followed by Rnase free water. You can also take chloroform-resistant plastic ware rinsed with chloroform to inactivate RNases.
- All buffers must be prepared with RNase-free ddH_2O
- When working with chemicals, always wear a suitable lab coat, disposable gloves and protective goggles.
- Change gloves frequently and keep tubes closed.
- All centrifugation steps are carried out at room temperature.
- To avoid cross contamination cavity seams should not be moisten with fluid.
- Reduce the preparation time as much as possible.
- Use only sterile, disposable polypropylene tubes throughout the procedure (these tubes are generally RNase free).
- Keep isolated RNA on ice.
- Do not use kit components from other kits with the kit you are currently using, unless the lot numbers are identical.
- To minimize the risk of infections from potentially infectious material, we recommend working under laminar air-flow until the samples are lysed.

This kit should only be used by personnel trained in *in- vitro* diagnostic laboratory practice.

Storage of RNA

Purified RNA can be stored -80°C and is stable for months and years, e.g. precipitated and stored in 70% ethanol.

Ordering information

Product	Package size	Catalogue No.
Invisorb® Spin Virus RNA Mini Kit	250 preparations	1040300300

Related products

InviMag® Virus RNA Kit/ KF96	5 x 96 preparations	7443300200
InviMag® Virus DNA/RNA Mini Kit/ KFmL	300 preparations	2441150400
RTP® Virus DNA/RNA Mini Kit	250 preparations	1040100300
Invisorb® Spin Virus DNA Mini Kit	250 preparations	1040200300

Possible suppliers for Isopropanol:

Carl Roth
2-Propanol
Rotipuran >99.7%, p.a., ACS, ISO
Order no. 6752

Applichem
2-Propanol für die Molekularbiologie
Order no. A3928

Sigma
2-Propanol
Order no. 59304-1L-F

Possible suppliers for Centrifuges:

Eppendorf AG
22331 Hamburg, Germany
Phone: +49 (0) 40 53801 0
Fax: +49 (0) 40 53801 556
E-Mail: eppendorf@eppendorf.com
www.eppendorf.com

SIGMA Laborzentrifugen GmbH
37507 Osterode am Harz, Germany
Phone: +49-5522-5007-0
Fax: +49-5522-5007-12
E-Mail: info@sigma-zentrifugen.de
www.sigma-zentrifugen.de

INVITEK
Molecular

Invitek Molecular GmbH
Röbert-Rössle-Str. 10
13125 Berlin

Phone: +49 30 94 89 29 01
Fax: +49 30 94 89 29 09
info@invitek-molecular.com

www.invitek-molecular.com

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